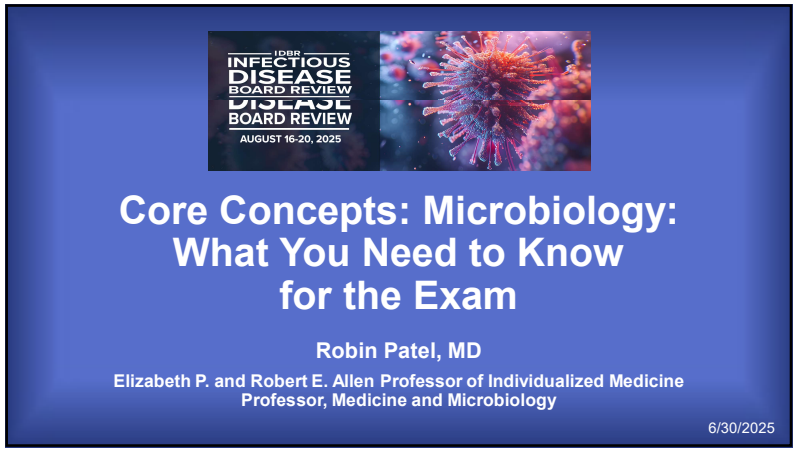
2 Core Concepts: Microbiology: What You Need to Know for the Exam

Speaker: Robin Patel, MD



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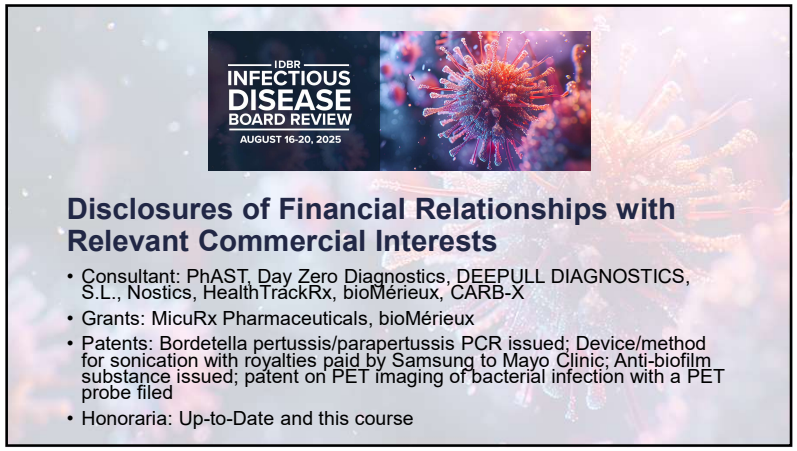

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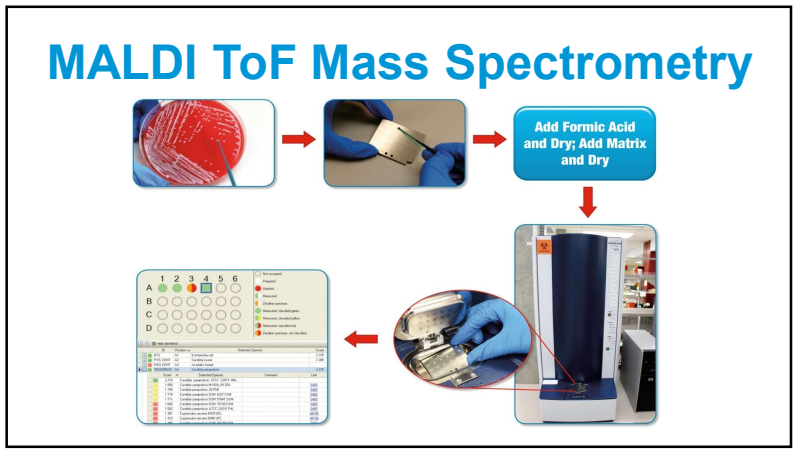

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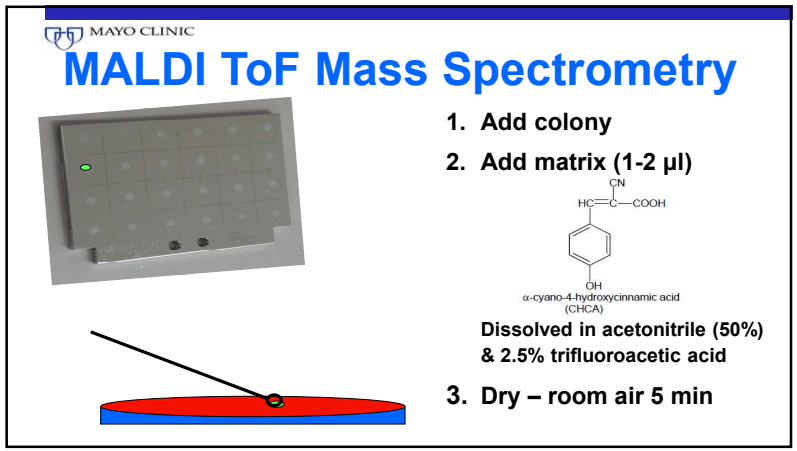

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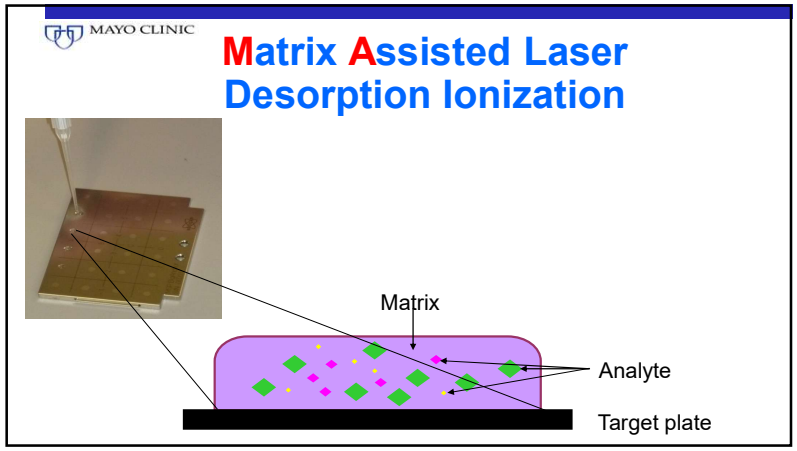

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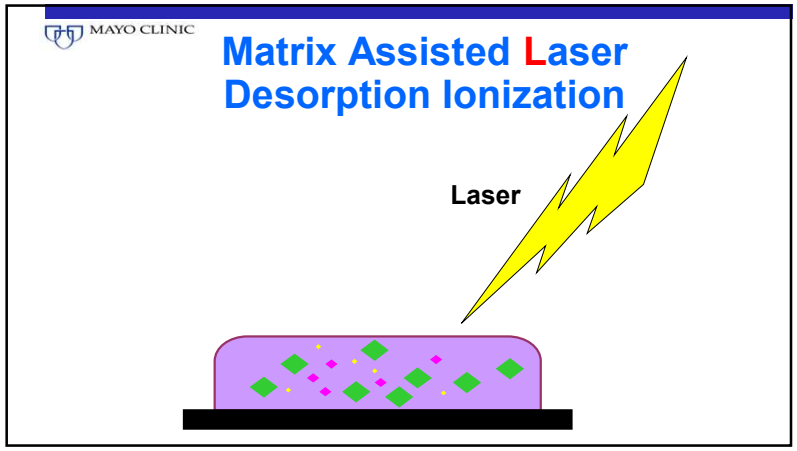

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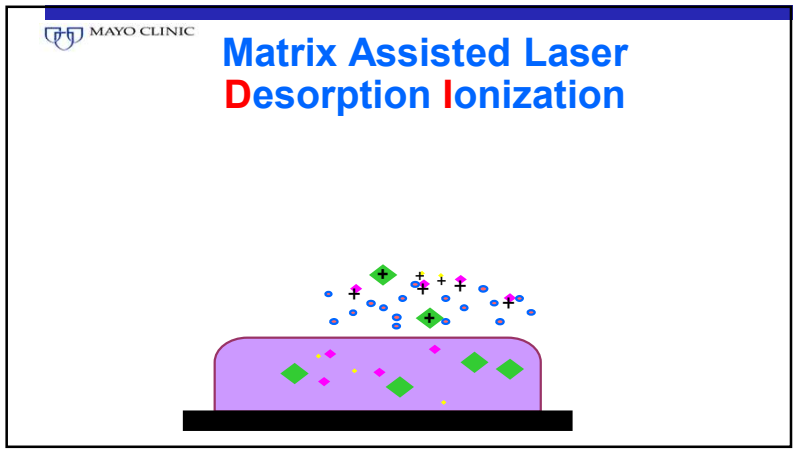

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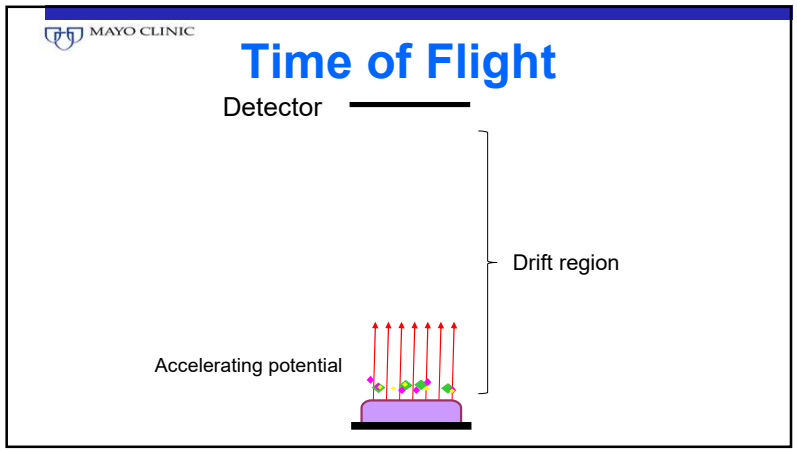

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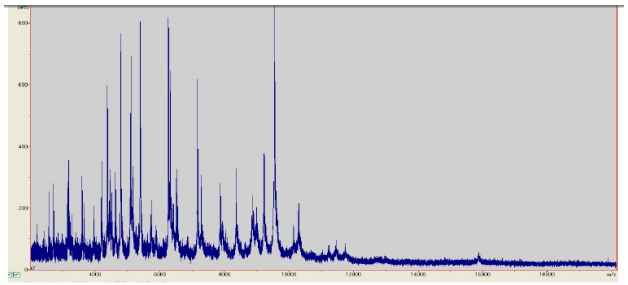
Mass Spectrum Generated Compared with Library (Database)



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Question #1

PREVIEW QUESTION

Which of the following will not grow on sheep blood, chocolate and/or MacConkey agar?

A. *Granulicatella adiacens*
B. *Bordetella pertussis*
C. *Brucella melitensis*
D. *Vibrio cholerae*
E. *Abiotrophia defectiva*

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Bacteria Requiring Specialized Media

- *Bordetella pertussis*
- *Brucella* species (+/-)
- *Burkholdheria pseudomallei* (+/-)
- *Campylobacter* species
- *Francisella tularensis* (+/-)
- *Helicobacter pylori*
- *Legionella* species
- *Mycoplasma* species (+/-)
- *Ureaplasma* species

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Question #2

Which of the following bacteria may stain acid-fast positive?

A. *Rhodococcus* species
B. *Cutibacterium* species
C. *Finegoldia* species
D. *Microbacterium* species
E. *Wolbachia* species

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Question #2

Which of the following bacteria may stain acid-fast positive?

A. *Rhodococcus* species*
B. *Cutibacterium* species
C. *Finegoldia* species
D. *Microbacterium* species
E. *Wolbachia* species

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Acid-fast Bacteria (Mycolic Acids)

- *Mycobacterium* species
- "Modified" acid fast stain positive
  - Weaker decolorizing agent (0.5-1% sulfuric acid in place of 3% acid-alcohol); do not stain well with Ziehl-Neelsen or Kinyoun stain
    - *Nocardia* species
    - *Rhodococcus* species
    - *Gordonia* species
    - *Tsukamurella* species
    - *Dietzia* species
- *Legionella micdadei* and some *Corynebacterium* species
  - [But not *Cutibacterium* species]

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Question #3

A laboratory technologist who has a longstanding history of diabetes mellitus inadvertently opens the lid of an agar plate growing an organism which is subsequently determined to be *Burkholdheria pseudomallei*.

You are asked to make a recommendation regarding postexposure prophylaxis.

Which of the following would you recommend?

A. Trimethoprim-sulfamethoxazole
B. Amoxicillin
C. Streptomycin
D. Cephalexin
E. None

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Which of the following would you recommend?

A. Trimethoprim-sulfamethoxazole *
B. Amoxicillin
C. Streptomycin
D. Cephalexin
E. None

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Burkholderia pseudomallei

- Postexposure antimicrobial prophylaxis
  - Trimethoprim-sulfamethoxazole
  - Doxycycline
  - Amoxicillin–clavulanic acid

Peacock SJ et al. Emerg Infect Dis. 2008 Jul http://wwwnc.cdc.gov/eid/article/14/7/07-1501

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Question #4

Which of the following, if present in a clinical specimen, poses a hazard for laboratory personnel?

A. Entamoeba histolytica
B. Trichuris trichiura
C. Enterobius vermicularis
D. Strongyloides stercoralis
E. Babesia microti

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Question #4

Which of the following, if present in a clinical specimen, poses a hazard for laboratory personnel?

A. Entamoeba histolytica
B. Trichuris trichiura
C. Enterobius vermicularis
D. Strongyloides stercoralis*
E. Babesia microti

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Strongyloides stercoralis

- Larvae - two forms
  1. Rhabditiform (in stool)
  2. Filariform

     Infectious stage that develops in soil and occasionally in patient (leads to autoinfection and is hazardous to laboratory personnel)
- Larvae detected
  - Microscopically (top) or
  - By placing feces on plate and detecting migrating larvae where they leave a trail of bacterial colonies (bottom)

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Laboratory- Acquired Bacterial, Fungal and Parasitic Infections (Selected)

- *Bacillus anthracis*
- *Brucella* species
- *Burkholdheria pseudomallei*
  - (*Burkholdheria mallei*)
- *Coxiella burnetii*
- *Coccidioides immitis/posadasii* (*Blastomyces dermatitidis, Histoplasma capsulatum*)
- Dermatophytes
- Enteric pathogens
- *Francisella tularensis*
- *Mycobacterium tuberculosis*
- *Neisseria meningitidis*
- *Salmonella enterica* subsp. *enterica* serovar Typhi
- *Staphylococcus aureus*
- *Strongyloides stercoralis*
- *Yersinia pestis*

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Organisms about which the Laboratory Should be Notified if Suspected

- Avian Influenza
- *Bacillus Anthracis*
- *Brucella* Species
- *Burkholdheria Pseudomallei*
- *Burkholdheria Mallei*
- *Clostridium Botulinum*
- *Coxiella Burnetii*
- *Coccidioides Immitis/Posadasii*
- Hemorrhagic Fever Viruses

  (e.g.., Ebola, Marburg, Chapare, Crimean-congo, Guanarito, Hanta, Junin, Kayasnur Forest Disease, Lassa Fever, Lujo, Machupo, Omsk Hemorrhagic Fever, Sabia)
- *Francisella Tularensis*
- Measles
- MERS, Sars-cov
- Nipah Virus, Hendra Virus
- Smallpox
- *Yersinia Pestis*

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FDA-Approved/Cleared Multiplex Panels for Gastrointestinal Pathogens in Stool (for reference)

	Verigene EP	xTAG® GPP	BioFire GIP	BioCode®	Qiastat-DX
Campylobacter species	✓	✓	✓	✓	✓
Salmonella species	✓	✓	✓	✓	✓
Shigella species/Enteroinvasive *E. coli*	✓	✓	✓	✓	✓
Vibrio species	✓		✓	✓	
Vibrio vulnificus					✓
Vibrio parahemolyticus				✓	✓
Vibrio cholerae		✓	✓		✓
Yersinia enterocolitica	✓	✓	✓	✓	✓
Escherichia coli 0157		✓	✓	✓	✓
Enterotoxigenic *E. coli*		✓	✓	✓	✓
Enteropathogenic *E. coli*			✓		✓
Enteroaggregative *E. coli*			✓	✓	✓
Plesiomonas shigelloides			✓		✓
Shiga toxin-producing *E. coli*	✓	✓	✓	✓	✓
Clostridioides difficile		✓	✓	✓	✓
Norovirus	✓	✓	✓	✓	✓
Rotavirus A	✓	✓	✓	✓	✓
Astrovirus			✓		✓
Adenovirus 40/41		✓	✓	✓	✓
Sapovirus			✓		✓
Cryptosporidium species		✓	✓	✓	✓
Entamoeba histolytica		✓	✓	✓	✓
Giardia lamblia		✓	✓	✓	✓
Cyclospora cayetanensis			✓		✓

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Gastroenteritis Panel Testing Key Points

- If available, culture independent methods of diagnosis recommended
- Indications: dysentery, moderate-to-severe disease, and symptoms lasting >7 days (define etiology, inform potential treatment)
- Not recommended for chronic diarrhea
- If C. *difficile* main consideration, test for *C. difficile* alone
- *Aerococcus* species not included

Riddle Et Al. Am J Gastroenterol 2016;111:602–622

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BIOFIRE FilmArray Meningitis/Encephalitis Panel (for reference)

Viruses	Bacteria	Fungi
Cytomegalovirus	*Escherichia coli* K1	*Cryptococcus neoformans/gattii*
Enterovirus	*Haemophilus influenzae*	
Herpes simplex virus 1	*Listeria monocytogenes*	
Herpes simplex virus 2	*Neisseria meningitidis*	
Human herpes virus 6	*Streptococcus agalactiae*	
Human parechovirus	*Streptococcus pneumoniae*	
Varicella zoster virus		

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Meningitis/Encephalitis Panel Key Points

- Doesn't nullify need for cell count, differential, protein, glucose, Gram stain, culture
- Cryptococcal antigen more sensitive than PCR
- *Streptococcus pneumoniae* antigen plus HSV, enterovirus and possibly VZV PCR an alternative
- May be helpful with current/recent antibiotic treatment
- HHV-6 & CMV may not be clinically significant

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Lower Respiratory Tract Panels (for reference)

	Curetis Unyvero	BioFire
Bacteria		
Acinetobacter spp.	✓	
Acinetobacter calcoaceticus-baumannii complex		✓
Chlamydia pneumoniae	✓	✓
Citrobacter freundii	✓	
Klebsiella aerogenes		✓
Enterobacter cloacae complex	✓	✓
Escherichia coli	✓	✓
Haemophilus influenzae	✓	✓
Klebsiella oxytoca	✓	✓
Klebsiella pneumoniae	✓	
Klebsiella pneumoniae group		✓
Klebsiella variicola	✓	
Legionella pneumophila	✓	✓
Moraxella catarrhalis	✓	✓
Morganella morganii	✓	
Mycoplasma pneumoniae	✓	✓
Proteus spp.	✓	✓
Pseudomonas aeruginosa	✓	✓
Serratia marcescens	✓	✓
Staphylococcus aureus	✓	✓
Stenotrophomonas maltophilia	✓	
Streptococcus agalactiae		✓
Streptococcus pneumoniae	✓	✓
Streptococcus pyogenes		✓

	Curetis Unyvero	BioFire
Viruses		
Influenza A		✓
Influenza B		✓
Respiratory Syncytial Virus		✓
Human Rhinovirus/Enterovirus		✓
Human Metapneumovirus		✓
Parainfluenza virus		✓
Adenovirus		✓
Coronavirus (non-SARS-CoV)		✓
Fungi		
Pneumocystis jirovecii	✓	
Resistance genes		
bla_{KPC}	✓	✓
bla_{NDM}	✓	✓
bla_{IMP}		✓
bla_{OXA-23}	✓	
bla_{OXA-24}	✓	
bla_{OXA-48}	✓	
bla_{OXA-58}		
$bla_{OXA-48-like}$		✓
bla_{VIM}	✓	✓
bla_{CTX-M}	✓	✓
bla_{TEM}	✓	
mecA	✓	
mecA/C and *MREJ*		✓

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Question #5

You are asked to see a 62-year-old man with a positive blood culture to advise on management.

Gram stain of the positive blood culture bottle shows Gram positive cocci in clusters.

A rapid PCR panel performed on the positive blood culture bottle contents detects *Staphylococcus aureus*, *Staphylococcus epidermidis* as well as *mecA/C* but not *mecA/C* and MREJ.

Which of the following is the interpretation of this finding?

A. Methicillin-susceptible *S. aureus* and methicillin-resistant *S. epidermidis*
B. Methicillin-susceptible *S. aureus* and methicillin-susceptible *S. epidermidis*
C. Methicillin-resistant *S. aureus* and methicillin-resistant *S. epidermidis*
D. Methicillin-resistant *S. aureus* and methicillin-susceptible *S. epidermidis*

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C. Methicillin-resistant *S. aureus* and methicillin-resistant *S. epidermidis*
D. Methicillin-resistant *S. aureus* and methicillin-susceptible *S. epidermidis*

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FDA-Approved Multiplex Panels for Detection of Gram-Positive Bacteria in Positive Blood Cultures (for reference)

	FilmArray MDx-Chex BCID2	VERIGENE® Gram-Positive Blood Culture Test	cobas® eplex BCID-GP Panel	cobas® eplex BCID-GN Panel
Staphylococcus species	✓	✓	✓	
Staphylococcus aureus	✓	✓	✓	
Staphylococcus epidermidis	✓	✓	✓	
Staphylococcus lugdunensis	✓	✓	✓	
Streptococcus species	✓	✓	✓	
Streptococcus agalactiae	✓	✓	✓	
Streptococcus pyogenes	✓	✓	✓	
Streptococcus pneumoniae	✓	✓	✓	
Streptococcus anginosus group		✓	✓	
Enterococcus species			✓	
Enterococcus faecalis	✓		✓	
Enterococcus faecium	✓	✓	✓	
Listeria species		✓	✓	
Listeria monocytogenes	✓		✓	
Bacillus cereus group			✓	
Bacillus subtilis group			✓	
Corynebacterium species			✓	
Cutibacterium acnes			✓	
Lactobacillus species			✓	
Micrococcus species		✓	✓	
Pan Gram-Positive				✓

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FDA-Approved Multiplex Panels for Detection of Gram-Negative Bacteria in Positive Blood Cultures (for reference), cont.

	FilmArray MDx-Chex BCID2	VERIGENE® Gram-Negative Blood Culture Test	cobas® eplex BCID-GP Panel	cobas® eplex BCID-GN Panel
Klebsiella oxytoca	✓	✓		✓
Klebsiella pneumoniae		✓		
Klebsiella pneumoniae group	✓			✓
Klebsiella aerogenes	✓	*		*
Salmonella species	✓			✓
Morganella morganii				✓
Stenotrophomonas maltophilia	✓			✓
Serratia species				✓
Serratia marcescens	✓			✓
Proteus species	✓	✓		✓
Proteus mirabilis				✓
Acinetobacter species		✓		
Acinetobacter baumannii				✓
Acinetobacter calcoaceticus-baumannii complex	✓			
Hemophilus influenzae	✓			✓
Cronobacter sakazakii				✓
Neisseria meningitidis	✓			✓
Pseudomonas aeruginosa	✓	✓		✓
Enterobacterales	✓			
Escherichia coli	✓	✓		✓
Enterobacter species		✓		✓
Enterobacter cloacae complex	✓			✓
Citrobacter species		✓		✓
Bacteroides fragilis	✓			✓
Fusobacterium necrophorum				✓
Fusobacterium nucleatum				✓
Pan Gram-Negative			✓	

*Detected as *Enterobacter* species

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FDA-Approved Multiplex Panels for Detection of Select Resistance Genes in Positive Blood Cultures (for reference), cont.

	FilmArray MDx-Chex BCID2	VERIGENE®		cobas®	
		Gram-Positive Blood Culture Test	Gram-Negative Blood Culture Test	eplex BCID-GP Panel	eplex BCID-GN Panel
mecA		✓		✓	
mecC				✓	
mecA/C	✓				
mecA/C and MREJ	✓				
vanA		✓		✓	
vanB		✓		✓	
vanA/B	✓				
***bla*$_{KPC}$**	✓		✓		✓
***bla*$_{NDM}$**	✓		✓		✓
***bla*$_{OXA}$**	✓		✓		✓
***bla*$_{VIM}$**	✓		✓		✓
***bla*$_{IMP}$**	✓		✓		✓
***bla*$_{CTX-M}$**	✓		✓		✓
mcr-1	✓				

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FDA-Approved Multiplex Panels for Detection of Fungi in Positive Blood Cultures (for reference), cont.

	FilmArray MDx-Chex BCID2	cobas®		
		ePlex BCID-GP Panel	eplex BCID-FP Panel	eplex BCID-GN Panel
Candida albicans	✓		✓	
Candida auris	✓		✓	
Candida dubliniensis			✓	
Candida famata			✓	
Nakaseomyces glabrata	✓		✓	
Candida guilliermondii			✓	
Candida kefyr			✓	
Pichia kudriavzevii	✓		✓	
Candida lusitaniae			✓	
Candida parapsilosis	✓		✓	
Candida tropicalis	✓		✓	
Cryptococcus gattii			✓	
Cryptococcus neoformans			✓	
C. neoformans/gattii	✓			
***Fusarium* species**			✓	
***Rhodotorula* species**			✓	
Pan *Candida*		✓		✓

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Staphylococci Methicillin Resistance

- Methicillin resistance mediated by *mecA* (or rarely *mecC*) gene products
  - Penicillin binding protein (PBP) target altered (PBP2a)
    - Confers resistance to all available β-lactams (except ceftaroline, ceftobiprole)
    - Even if staphylococci that are methicillin-resistant *appear* susceptible to these other β-lactams, they are not effective
- Oxacillin or cefoxitin tested
- *mecA/C* and MREJ specific for *Staphylococcus aureus*
- For serious infections, oxacillin susceptibility confirmed using PBP2a testing or *mecA* (and *mecC*) nucleic acid amplification test (NAAT)

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FDA-Approved Rapid Phenotypic Susceptibility Tests - Positive Blood Culture Bottles

- Accelerate Diagnostics
- Selux$_{DX}$ Next-Generation Phenotyping (NGP) System
- VITEK® REVEAL™ AST System
- Q-linea ASTar system
- Lifescale

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BioFire Joint Infection Panel (Synovial Fluid)

Anaerococcus prevotii/vaginalis	*Escherichia coli*
Clostridium perfringens	*Haemophilus influenzae*
Cutibacterium avidum/granulosum	*Kingella kingae*
Enterococcus faecalis	*Klebsiella aerogenes*
Enterococcus faecium	*Klebsiella pneumoniae* group
Finegoldia magna	*Morganella morganii*
Parvimonas micra	*Neisseria gonorrhoeae*
Peptoniphilus	*Proteus* spp.
Peptostreptococcus anaerobius	*Pseudomonas aeruginosa*
Staphylococcus aureus	*Salmonella* spp.
Staphylococcus lugdunensis	*Serratia marcescens*
Streptococcus species	*Candida* spp.
Streptococcus agalactiae	*Candida albicans*
Streptococcus pneumoniae	bla_{IMP}, bla_{KPC}, bla_{NDM}, $bla_{OXA-48-like}$,
Streptococcus pyogenes	bla_{VIM}, bla_{CTX-M}
Bacteroides fragilis	*mecA/C* and MREJ
Citrobacter	*vanA/B*
Enterobacter cloacae complex	

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Question #6

A 65-year-old man has multiple blood cultures positive for *Pseudomonas aeruginosa* resistant to amikacin, gentamicin, tobramycin, aztreonam, cefepime, ceftazidime, meropenem, piperacillin-tazobactam, ciprofloxacin, and levofloxacin. You call the clinical microbiology laboratory to request susceptibility testing of an additional antimicrobial.

Which of the following is most appropriate?

A. Dalbavancin
B. Tedizolid
C. Ceftolozane/tazobactam
D. Oritavancin

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B. Tedizolid
C. Ceftolozane/tazobactam*
D. Oritavancin

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Question #7

PREVIEW QUESTION

You are asked to see a 43-year-old woman to advise on management of a positive blood culture.

- Gram stain of her blood culture bottle shows Gram-negative bacilli.
- A rapid PCR panel performed on the positive blood culture bottle contents detects *Klebsiella pneumoniae* and bla_{KPC}.

The bla_{KPC} gene product would be expected to confer resistance to which of the following?

A. Cefepime
B. Plazomicin
C. Colistin
D. Ceftazidime/avibactam

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IDBR INFECTIOUS DISEASE BOARD REVIEW 2025

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B. Plazomicin
C. Colistin
D. Ceftazidime/avibactam

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Typical Susceptibility of a bla_{KPC}-Producer

Klebsiella pneumoniae

Ampicillin	>16 R	**Ampicillin/Sulbactam**	>16/8 R	**Piperacillin/Tazobactam**	64/4 R
Cefazolin	>16 R	**Oral cephalosporins**	R	**Cefepime**	>16 R
Ceftazidime	>16 R	**Ceftriaxone**	>32 R	**Ertapenem**	>1 R
Meropenem	>8 R	**Aztreonam**	>16 R	**Ciprofloxacin**	>2 R
Levofloxacin	4 I	**Amikacin**	>32 R	**Gentamicin**	>8 R
Tobramycin	4 S	**Tigecycline**	2 S	**TMP/SMX**	>2/38 R

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Typical Susceptibility of an ESBL-producer

Escherichia coli

Ampicillin	>16 R	**Ampicillin/Sulbactam**	>16/8 R	**Piperacillin/Tazobactam**	S/R*
Cefazolin	>16 R	**Oral cephalosporins**	R	**Cefepime**	S/SDD/R
Ceftazidime	>16 R	**Ceftriaxone**	>32 R	**Ertapenem**	≤0.5 S
Meropenem	≤1 S	**Aztreonam**	>16 R	**Ciprofloxacin**	≤1 S
Levofloxacin	≤2 S	**Amikacin**	≤8 S	**Gentamicin**	≤1 S
Tobramycin	4 S	**Tigecycline**	2 S	**TMP/SMX**	>2/38 R

*Not currently recommended for infection outside of urinary tract

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Typical Susceptibility of Inducible, Chromosomally-Encoded AmpC β-Lactamase Producer

*Enterobacter cloacae**

Ampicillin	>16 R	**Ampicillin/Sulbactam**	>16/8 R	**Piperacillin/Tazobactam**	S/R*
Cefazolin	>16 R	**Oral cephalosporins**	R	**Cefepime**	S/SDD
Ceftazidime	>16 R	**Ceftriaxone**	>32 R**	**Ertapenem**	≤0.5 S
Meropenem	≤1 S	**Aztreonam**	S/R	**Ciprofloxacin**	≤1 S
Levofloxacin	≤2 S	**Amikacin**	≤8 S	**Gentamicin**	≤1 S
Tobramycin	4 S	**Tigecycline**	2 S	**TMP/SMX**	>2/38 R

Enterobacter cloacae, Klebsiella aerogenes, Citrobacter freundii
****Avoid ceftriaxone or ceftazidime even if test susceptible; cefepime an acceptable choice**
IDSA Guidance on the Treatment of Antimicrobial-Resistant Gram-Negative Infections (idsociety.org)

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Question #8

Which of the following susceptibility patterns would be typical for an *Escherichia coli* isolate carrying a New Delhi metallo-β-lactamase (NDM)?

	Cefazolin	Cefotaxime	Ceftazidime	Piperacillin/ tazobactam	Imipenem	Aztreonam
A.	R	S	S	S	S	S
B.	R	R	R	S	S	R
C.	R	R	R	R	S	R
D.	R	R	R	R	R	R

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	Cefazolin	Cefotaxime	Ceftazidime	Piperacillin/ tazobactam	Imipenem	Aztreonam
A.	R	S	S	S	S	S
B.	R	R	R	S	S	R
C.	R	R	R	R	S	R
D.*	**R**	**R**	**R**	**R**	**R**	**R**

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Question #9

Which of the following tests for carbapenemase production?

A. PBP2a test
B. D-test
C. Carba NP test
D. Polymerase chain reaction assay

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Question #9

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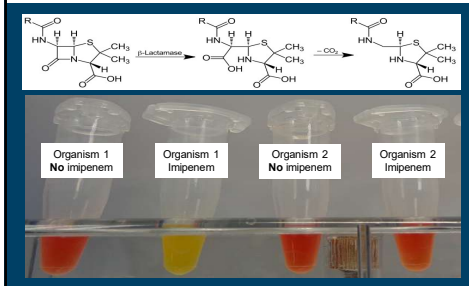
Carbapenemase Production Test
Carba NP Test



- β-lactam ring hydrolyzed by carbapenemase
- ↓pH (detected by indicator dye color change red → yellow)
- Rapid (2 hours)

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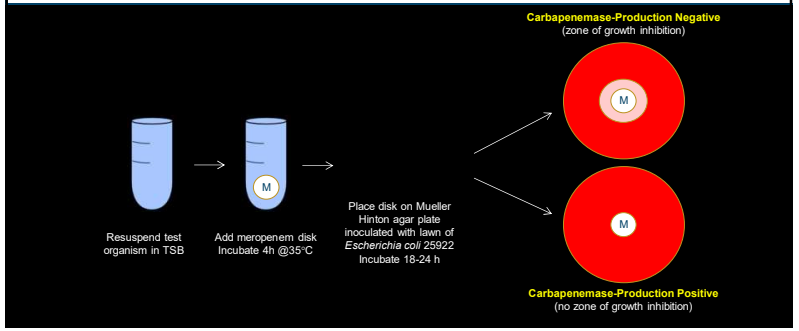
Carbapenemase Production Test
Modified Carbapenem Inactivation

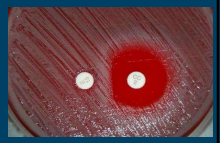


50

Question #10

The image shows *Staphylococcus aureus* grown with an erythromycin disc (left) and a clindamycin disc (right).

Which of the following is the correct interpretation of these results?



A. Erythromycin susceptibility, inducible clindamycin resistance
B. Erythromycin resistance, constitutive clindamycin resistance
C. Erythromycin resistance, inducible clindamycin resistance
D. Erythromycin susceptibility, constitutive clindamycin resistance

51

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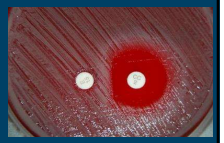

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C. Erythromycin resistance, inducible clindamycin resistance*
D. Erythromycin susceptibility, constitutive clindamycin resistance

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Inducible Clindamycin Resistance (D-Test)

- Macrolide resistance from alteration in ribosomal target → co-resistance to clindamycin; constitutive or inducible
  - Constitutive, erythromycin & clindamycin test resistant
  - Inducible, erythromycin tests resistant but clindamycin tests falsely susceptible
- (Macrolide resistance due to efflux → no effect on clindamycin)

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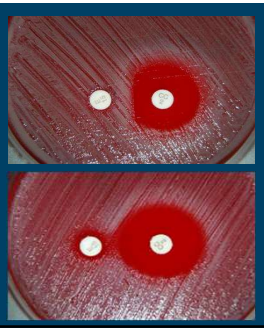
Inducible Clindamycin Resistance (D-Test)

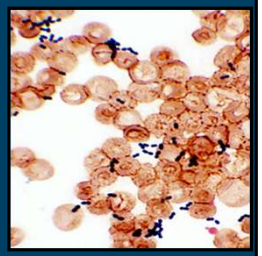
- Erythromycin & clindamycin disks incubated on plate
  - Flattening of zone of inhibited growth between disks = inducible clindamycin resistance (top)
  - If erythromycin does not influence zone around clindamycin disk, clindamycin susceptible (bottom)



54

Question #11

- You are asked to see a 95-year-old woman who is a resident of a long-term care facility to advise on therapy for bacteremia associated with a urinary tract infection.
- She has had two sets of blood cultures collected, both of which signaled positive after 17 hours of incubation.
- Gram stain of the bottles is shown.
- A rapid PCR panel performed on the positive blood culture bottle detects *Enterococcus* species as well as *vanA/vanB*.



55

Question #11

Which of the following is the most likely identity of the blood culture isolate?

A. *Enterococcus gallinarum*
B. *Enterococcus faecium*
C. *Enterococcus faecalis*
D. *Enterococcus casseliflavus*
E. *Enterococcus avium*

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B. *Enterococcus faecium**
C. *Enterococcus faecalis*
D. *Enterococcus casseliflavus*
E. *Enterococcus avium*

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Enterococci
Vancomycin Susceptibility Testing

- Vancomycin MICs ≥32 µg/ml
  - Typically, VanA or VanB mediated resistance
  - Typically, *E. faecium*
  - Epidemiologically significant
- Vancomycin MICs, 8-16 µg/ml (intermediate)
  - VanC
  - *E. gallinarum* or *E. casseliflavus/flavescens*
  - Not epidemiologically significant

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Question #12

PREVIEW QUESTION

A 44-year-old man who underwent bilateral lung transplantation for pulmonary hypertension develops a sternal wound infection with sternal dehiscence 15 days post-transplant.

Blood cultures are negative. He undergoes sternal debridement with the finding of purulence and negative Gram and KOH stains.

After three days of incubation, pinpoint, clear colonies are visualized on cultures on sheep blood agar, however Gram stain of these colonies is negative.

Which of the following is the most appropriate empiric antibiotic to treat this patient?

A. Cefepime
B. Ceftriaxone
C. Trimethoprim-sulfamethoxazole
D. Azithromycin
E. Doxycycline

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C. Trimethoprim-sulfamethoxazole
D. Azithromycin
E. Doxycycline *

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Mycoplasma hominis

- Post-cardiothoracic transplant
  - Pleuritis, surgical site infection and/or mediastinitis
- Treatment
  - Inactive
    - Cell wall active antibiotics
    - Trimethoprim/sulfamethoxazole
    - Aminoglycosides
    - Erythromycin and azithromycin
  - Active
    - Tetracyclines (doxycycline preferred)
    - Fluoroquinolones
    - Clindamycin

Sampath, R., et al. EBioMedicine (2017), http://dx.doi.org/10.1016/j.ebiom.2017.04.026

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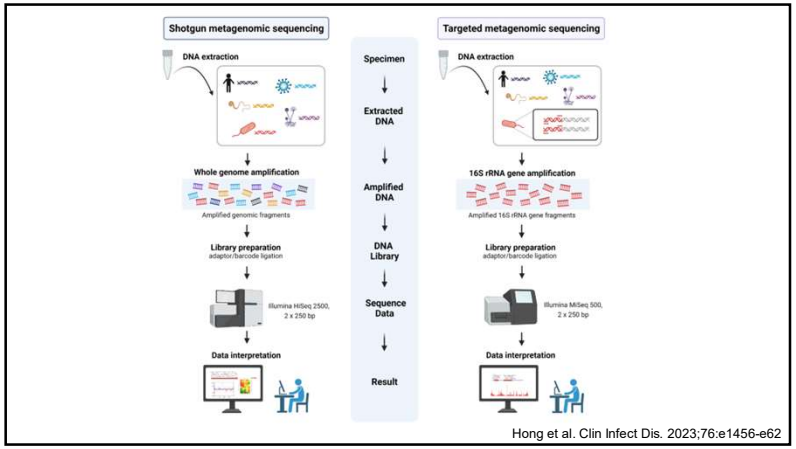

Hong et al. Clin Infect Dis. 2023;76:e1456-e62

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2023 Duke-International Society For Cardiovascular Infectious Diseases Criteria For Infective Endocarditis (IE)

- Pathologic Criteria - Microorganisms detected (appropriate sample) - PCR, amplicon/metagenomic sequencing, *in situ* hybridization
- Blood cultures - Removed required timing (and separation) venipunctures for blood cultures
- MAJOR CRITERIA
  - Positive blood cultures - Microorganisms that commonly cause IE ≥2 blood culture sets (typical) or that occasionally or rarely cause IE ≥3 blood culture sets (nontypical)
    - Typical: *Staphylococcus aureus*; HACEK group; *Staphylococcus lugdunensis*; *Enterococcus faecalis*; all streptococcal species (except for *Streptococcus pneumoniae* and *pyogenes*), *Granulicatella* and *Abiotrophia* spp., *Gemella* spp., In setting of intracardiac prosthetic material, include these as "typical" pathogens: coagulase negative staphylococci, *Corynebacterium striatum* and *jeikeium*, *Serratia marcescens*, *Pseudomonas aeruginosa*, *Cutibacterium acnes*, nontuberculous mycobacteria (especially *M. chimaerae*), *Candida* spp.
- MAJOR CRITERIA
  - Blood PCR or amplicon/metagenomic sequencing detection of *Coxiella burnetii*, *Bartonella* spp., *Tropheryma whipplei*
  - IFA ≥1:800 for IgG antibodies *Bartonella henselae* or *B. quintana*
- MINOR CRITERIA - Positive culture, PCR, or other nucleic acid-based test (amplicon/metagenomic sequencing, *in situ* hybridization) organism consistent with IE from sterile body site other than cardiac tissue, cardiac prosthesis, or arterial embolus; or single finding of skin bacterium by PCR on valve or wire without additional clinical or microbiological supporting evidence

Fowler et al. Clin Infect Dis. 2023;77:518-26.

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Shotgun Metagenomic Sequencing - CSF

- May identify CNS infection not found with conventional diagnostics
  - e.g., hepatitis E virus, St. Louis encephalitis virus, *Taenia solium*
- Not always positive in CNS infection
  - Infections for which serology preferred (e.g., syphilis, Lyme)
  - Low pathogen titer in CSF
  - Diagnosis from samples other than CSF (tissue)
- May detect microorganisms that are not cause of CNS infection
  - e.g., HHV-6, EBV

Wilson et al. N Engl J Med. 2019;380:2327-40

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Plasma Shotgun Metagenomic Sequencing (Cell Free DNA)

- 18,690 reports; 15,165 patients; 22,792 detections
- 701 unique taxa
  - 75% bacteria
  - 15% fungi
  - 7% viruses (herpesviruses, BK virus, adenovirus, torque teno virus)
  - 3% parasites

Park et al. J Clin Microbiol. 2023;61:e01855

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Plasma Shotgun Metagenomic Sequencing (Cell Free DNA)

- *Legionella*-like bacteria (80), 40% *Legionella pneumophila*
- *Nocardia* detections (76), 25% *Nocardia cyriacigeorgica*
- *Mycobacterium* species detections (156)
- Zoonotic and vector-borne bacteria detections (247)
  - *Bartonella henselae, Bartonella quintana, Bartonella clarridgeiae, Bartonella vinsonii*
  - *Rickettsia typhi, Rickettsia rickettsii, Rickettsia felis*
  - *Ehrlichia chaffeensis, Ehrlichia muris, Anaplasma phagocytophilum, Borrelia hermsii, Borrelia burgdorferi*
  - *Coxiella burnetii, Streptobacillus moniliformis, Capnocytophaga canimorsus*
  - *Leptospira kirschneri, Leptospira interrogans, Leptospira santarosai, Leptospira borgpetersenii, Leptospira noguchii*
  - *Francisella tularensis, Brucella melitensis*
- 632 candidal, 374 *Aspergillus* species, 196 Mucorales, 78 dimorphic and 33 dematiaceous fungal detections
- 9 microsporidia
- 57 protozoa - 68% *Toxoplasma gondii*, 14% pathogenic amoebae
- 7 helminths - 4 *Strongyloides stercoralis*, 2 *Echinococcus multilocularis*, 1 *Schistosoma mansoni*

Park et al. J Clin Microbiol. 2023;61:e01855

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Question #13

You are consulted to advise on the course of action for a 57-year-old female liver transplant recipient (transplant for alcoholic steatohepatitis; CMV D^+/R^+) who has a whole blood HHV-6 viral load of $3.6x10^6$ copies/ml at three months post-transplant. The test was performed because of a report of subjective fever of four days' duration. She has no other new symptoms. The patient received one month of acyclovir prophylaxis post-transplant and is currently receiving mycophenolate mofetil, prednisone and trimethoprim-sulfamethoxazole. Her post-transplant course was complicated by one episode of treated rejection on day 30 post transplant. Physical examination is unremarkable, and she is afebrile.

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Question #13

Which of the following would you recommend?

A. Intravenous ganciclovir
B. Oral valganciclovir
C. Oral acyclovir
D. Intravenous foscarnet
E. No antiviral therapy is indicated

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Question #13

Which of the following would you recommend?

A. Intravenous ganciclovir
B. Oral valganciclovir
C. Oral acyclovir
D. Intravenous foscarnet
E. No antiviral therapy is indicated*

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Chromosomally Integrated Human Herpesvirus-6

- High HHV-6 levels in whole blood
  - (>5.5 $\log_{10}$ copies/ml)
  - Suggest chromosomally integrated HHV-6
- 1:1 ratio of viral to human genomes

Pellett et al. Rev Med Virol 2012;22:144-55

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Question #14

A 76-year-old woman presents with three days of cough, difficulty breathing and fever. She has never received a COVID-19 vaccine and has never been diagnosed with COVID-19.

Which of the following COVID-19 tests is recommended?

A. Antigen
B. Serology
C. NAAT

71

Question #14

A 76-year-old woman presents with three days of cough, difficulty breathing and fever. She has never received a COVID-19 vaccine and has never been diagnosed with COVID-19.

Which of the following COVID-19 tests is recommended?

A. Antigen
B. Serology
C. NAAT*

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Covid-19 Diagnostics

- NAAT generally preferred over antigen testing
  - Symptomatic individuals suspected of having COVID-19
  - Asymptomatic individuals exposed to SARS-CoV-2 infection
  - Interpret Ct values with caution
- Healthcare provider or patient collected specimens acceptable
- Swabs from nasopharynx, anterior nares, oropharynx, or mid-turbinate regions; saliva or mouth gargle acceptable
  - Compared to nasopharyngeal swabs, anterior nares or oropharynx swabs alone yield more false-negative results than combined anterior nares/oropharynx swabs, mid-turbinate swabs, saliva, or mouth gargle
- Suspected lower respiratory infection → upper respiratory sample; if negative, lower respiratory sample

IDSA Guidelines on the Diagnosis of COVID-19

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Question #15

A 22-year-old man presents to an urgent care clinic with a cough of 9 day's duration. Prior to the onset of cough, he experienced a runny nose for a couple of days. He has not had fever. The cough is keeping him up at night and has caused him to vomit on two occasions. He has not received any vaccinations since he was a baby. Physical examination is unremarkable, except that he looks tired.

Which of the following is the test of choice for this patient?

A. Nasopharyngeal swab antigen
B. Nasopharyngeal swab culture
C. Nasopharyngeal swab NAAT
D. Serology

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Question #15

A 22-year-old man presents to an urgent care clinic with a cough of 9 day's duration. Prior to the onset of cough, he experienced a runny nose for a couple of days. He has not had fever. The cough is keeping him up at night and has caused him to vomit on two occasions. He has not received any vaccinations since he was a baby. Physical examination is unremarkable, except that he looks tired.

Which of the following is the test of choice for this patient?

A. Nasopharyngeal swab antigen
B. Nasopharyngeal swab culture
C. Nasopharyngeal swab NAAT*
D. Serology

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Question #16

A 60-year-old man presents to an emergency department in Texas with a maculopapular rash that began 7 days prior to presentation. The rash started on his face and subsequently spread to his neck, chest and then arms and legs, and has been accompanied by fever. Before onset of the rash, he had two days of rhinorrhea and red eyes.

Which of the following is recommended?

A. Throat or nasopharyngeal swab NAAT and serum IgM test
B. Serum IgM test
C. Urine NAAT and serum IgG test
D. Throat or nasopharyngeal swab culture and serum IgM test

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B. Serum IgM test
C. Urine NAAT and serum IgG test
D. Throat or nasopharyngeal swab culture and serum IgM test

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